

Intended Use

For **IN VITRO quantitative** determination of Triglycerides in serum.

Clinical Significance

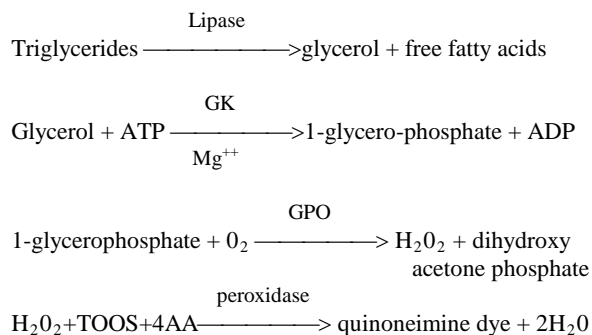
Determination of Triglycerides are primarily used for diagnosing atherosclerosis and heart disease, diabetes mellitus, nephritis, biliary obstruction and metabolic disorders associated with endocrine disturbances, as well as for monitoring the causes and treatments.

Method History

Various methods for the determination of Triglycerides in human serum are currently in use in the routine clinical laboratory. With the advent of stable, more specific lipolytic enzymes, simpler and more efficient procedures have been described. (1,2) The Catachem Triglycerides method described below uses glycerophosphate oxidase coupled with a sensitive colorimetric endpoint reaction based on the work of Trinder. (3)

Method Principle

Serum Triglycerides are hydrolyzed by microbial lipase to glycerol and free fatty acids. The resultant glycerol, in the presence of glycerol kinase (GK) ATP and Mg^{++} ions is phosphorylated to glycerol-1-phosphate. This latter metabolite is then oxidized by glycerophosphate oxidase (GPO) to produce hydrogen peroxide. The hydrogen peroxide thus produced is quantitatively determined by coupling 4-aminoantipyrine with N-ethyl-N-(2-hydroxy-3-sulphopropyl)-m-toluidine (TOOS) (3-4), where a quinonemine dye with maximum absorption at 550 nm is produced. The following reaction scheme illustrates the reactions that occur in this method:



Reagent Content

The concentration of the active ingredients in the reagent will be approximately as follows:

Triglycerides GPO-Trinder Reagent

One liter **R1** contains:

Buffer	
ATP	0.30 mM
4-aminoantipyrine	0.15 mM
Lipase	1000 U
Glycerol Kinase	1000 U
Glycerophosphate oxidase	2500 U
Peroxidase	2000 U

One liter **R2** contains:

TOOS	7.2 mM
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Precautions

Avoid contact of the reagent with skin and eyes. Should contact occur, wash affected area with plenty of cold water.

DO NOT PIPETTE REAGENTS BY MOUTH

Preparation Of Working Reagent

Catachem Triglycerides Reagent is packaged as a liquid R1 component and liquid R2 component. No preparation is required.

Reagent Storage And Stability

Store the reagents at 2-8°C. When stored as directed, these reagents are stable until the expiration date stated on the label. Upon opening and if stored capped when not in use, each TG Reagent component is stable for 60 days at 2-8°C.

Specimen Collection And Preparation

The use of clear, unhemolyzed serum is recommended. A fasting specimen is necessary for an accurate triglycerides determination.

Interfering Substances

A number of substances have been reported to affect the accuracy of triglycerides methods using the oxidase-peroxidase procedures (3,2). A summary of the influence of drugs on clinical laboratory procedures may be found by consulting D.S. Young, et al (5).

Expected Values

The "risk values" for Triglycerides in a human population as defined by the National Heart and Lung Institute are listed below. Levels in animals differ species to species.

AGE (Years)	TRIGLYCERIDES (mg/dL)
20-29	140
30-49	160
50-59	190

It is recommended that each laboratory establish the normal ranges for the species being tested and the species location.

$$\begin{aligned} \text{Calibrator (mg/dL)} &= 200 \\ &0.300 \\ \text{Triglycerides (mg/dL)} &= \frac{\text{-----}}{0.250} \times 200 = 240 \text{ mg/dL} \end{aligned}$$

Directions For Use

The Catachem Triglycerides Method requires one single reagent. Upon opening, the Working Reagent is stable for a minimum 60 days when stored capped at 2-8°C.

Procedure

Important: Read the entire instructions procedure before proceeding with the assay.

Materials Required But Not Provided

- Spectrophotometer
- Cuvettes 1 cm light path
- Timer to time incubation time
- Pipette 1.0 ml for reagent
- Pipette 0.010 ml for sample

Materials Provided

Catachem Triglycerides Reagent

Analytical Parameters

- Wavelength 550nm
- Temperature 37°C
- Pathlength 1 cm
- Reaction Mode Endpoint
- Reaction Time 5 min
- Reagent Volume R1 0.9 ml
- Reagent Volume R2 0.1 ml
- Sample Volume 0.01 ml (10µL)
- Total Volume 1.01 ml
- Sample-to-Reagent Ratio 1:100

Assay Procedure

1. Pipette 0.9 ml of Catachem Triglycerides Reagent R 1 into each of three cuvettes marked "Calibrator", "Sample", and "Blank".
2. Pipette 0.1 ml of Catachem Triglycerides Reagent R 2 into each of the same three cuvettes marked "Calibrator", "Sample", and "Blank"
2. Pipette 0.01 ml (10 uL) of Calibrator or Sample into their respective cuvettes. Mix all cuvettes well.
3. Incubate all cuvettes for 5 minutes at 37°C.
4. Set spectrophotometer wavelength at 550nm and zero the instrument with the cuvette marked "Blank".
5. Read the "Calibrator" and "Sample" absorbencies.
6. Calculate the Triglycerides concentration (mg/dL) in the Sample(s), as shown in "results and calculations" below.

Results And Calculations

$$\text{Triglycerides (mg/dL)} = \frac{\text{Sample Abs}}{\text{Calibrator Abs}} \times \text{Calibrator (mg/dL)}$$

Example:

$$\begin{aligned} \text{Sample absorbance} &= 0.300 \\ \text{Calibrator absorbance} &= 0.250 \end{aligned}$$

Quality Control

To monitor the performance of the Working Reagent and the procedure used, we recommend the regular use of a normal and abnormal control serum.

Method Performance Characteristics

Sensitivity: 0.0008-0012 absorbance units per mg/dL.

Linear Range: 0-1000 mg/dL.

Precision: Within-run and day-to-day precision is summarized below:

Triglycerides Precision Study

TGS	Within-Run		Total Precision	
Mean	SD	CV	SD	CV
mg/dL	mg/dL	%	mg/dL	%
69	1.1	1.6	1.5	2.1
386	1.2	0.3	2.9	0.7
714	3.5	0.5	7.4	1.0

Correlation

A comparison of the method using an automated analyzer and a reference method resulted in the following regression statistics:

$$\begin{aligned} \text{Range} &= 29-715 \text{ mg/dL} \\ \text{N} &= 118 \\ \text{Y} &= 1.036-5.8 \\ \text{r} &= 0.998 \\ \text{Sy.x} &= 7.0 \end{aligned}$$

References

1. Bucolo G, David, H. Clin Chem 20, 47648(1973).
2. Fossati P, Prencipe L. Clin Chem 28, 10 (1980).
3. McGowan MW, Artis JD, Strandbergh DR and Zak B. Clin Chem 29, 3 (1983).
- 5 .Trinder P, Ann Clin Biochem 6, 24 (1969).
4. Katsumi Tamaoku, Keiuy Ueno, Kayoko Akiura and Yosuke Ohkura. Chem PharmBull 30 (7) 2492-2497 (1982).
5. Young DS, Pestaner LD, Gibberman V. Clin Chem 21, 5 (1975).

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